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Current Advances in Regenerative Medicine for Articular Cartilage Injury: Progress and Market Trends

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Introduction

Articular cartilage is a complex organ with connective tissue that has a limited repair capacity [1]. Usually, small lesions that penetrate the subcutaneous bone layer are repaired by creating a fibrous scar, but extensive injuries require medical intervention. In recent years, a variety of surgical and non-surgical treatments have been developed to repair cartilage, but the complete treatment of lesions larger than 2-4 mm of the articular cartilage remains a therapeutic challenge [2].

Recently, tissue engineering and regenerative medicine using stem cells and biomaterials were able to revolutionize tissue and organ regeneration [3]. This method allows custom design for tissue regeneration and offers tissue replacement that mimics native tissue without adverse effects such as suppression

Abstract

Today, treatments of cartilage and osteochondral lesions are routine clinical procedures. Treatment of large Articular Cartilage (AC) defects is technically difficult and complex, often accompanied by failure. Articular cartilage is a highly specialized connective tissue with limited ability to repair itself after injury due to a lack of blood vessels, lymph, and nerves. Therefore, without sufficient and potent intervention, cartilage lesions can easily lead to progressive tissue degeneration, disabling joint pain, and eventually the degenerative disease, Osteoarthritis (OA). Various treatments for cartilage regeneration have shown encouraging results, but unfortunately, none of them have been the perfect solution. New minimally invasive and effective techniques are being developed. The development of tissue engineering technology has created strong promise to engineer or regenerate functional and healthy articular cartilage. In this technology, potential stem cell sources are mainly supplied with pluripotent stem cells and mesenchymal stem cells from various sources. In the meantime, some such as BioSeed®-C and NOVOCART® have been marketed. In this review, the common techniques of articular cartilage reconstruction and the clinical application of articular cartilage tissue engineering are described in detail.

Keywords: Articular cartilage; Cartilage gene therapy; Cartilage product; Regenerative medicine; Tissue engineering; Transplantation methods

of the immune system or contamination of the donor disease [4]. One of the major challenges in this method is designing appropriate scaffolds with the structure of native tissues [5]. Tissue-engineered cartilage must be highly compatible to prevent acute immune response after transplantation, also it must have special properties such as the ability to combine with subcutaneous bone and adjacent cartilage, mimic the mechanical properties of natural cartilage to maintain function, and withstand long-term loads [6].

In this study, new methods of repairing extensive joint cartilage injuries using different cellular sources and synthesis techniques are reviewed and a list of commercial products used in the treatment of cartilage injuries is presented.

Tissue Engineering Strategies for Articular Cartilage Regeneration

Current strategies for repairing articular cartilage, including surgical and non-surgical treatments, have not yet provided long-term solutions for repairing large articular cartilage lesions [2]. Tissue engineering and regenerative medicine can provide alternative treatment strategies using appropriate scaffolding, cells, and biochemical and biomechanical stimuli [3]. This method allows a custom design to regenerate native tissue without side effects such as infection transmission or the use of immunosuppressive drugs [5]. Tissue-engineered cartilage must be biocompatible and can connect with the subcutaneous bone and adjacent cartilage. In addition, it should be able to mimic the physical and mechanical properties of native tissue [6].

Scaffold for Articular Cartilage Tissue Engineering

In recent years, various scaffolds including synthetic or natural materials such as polylactides, polyglycolide, hyaluronic acid, collagen, and silk have been studied for articular cartilage tissue engineering [7]. In previous studies, a matrix derived from decellularized cartilage was used as a natural source for scaffolding in cartilage regeneration. This substrate was able to synthesize the extracellular matrix of cartilage by inhibiting the hypertrophic differentiation of embedded Mesenchymal Stem Cells (MSCs). The results also showed that the synthesized extracellular matrix could support the differentiation of mesenchymal stem cells into fibroblast and fibrochondrocyte phenotypes [8]. Hydrogels are another class of materials used as scaffolds in articular cartilage tissue engineering. These materials have received a lot of attention due to their injectability and ductility compatible with irregular defects of articular cartilage. On the other hand, due to the advent of 3D printing technology, achieving the right hydrogel can be a big step in the design and customization of graft implants in the repair of cartilage defects. In addition, in this technology, cells and growth factors can be included in the scaffold structure during synthesis [9]. In previous research has used synthetic polymers such as polycaprolactone and polylactic acid, as well as natural sources such as alginate and hyaluronan to create custom anatomical scaffolds for articular cartilage in 3D printers [10]. Stimuli-responsive hydrogels or smart hydrogels are a group of hydrogels in which a specific transition occurs due to small changes in the environment [11]. This group of hydrogels in response to various external physicochemical factors such as chemical stimuli [12], temperature changes [13], solvent type [14], pH [15], ionic strength [16], wavelength or light intensity [17] or electric fields and magnetically are sensitive [18].

The use of smart hydrogels in actuators, sensors, scaffolds, and drug delivery has received considerable attention because of their rapid response to environmental stimuli, which can cause significant changes. Designing scaffolds based on intelligent injection hydrogels with nanostructured properties and rapid response to stimuli can be an appropriate option to meet all the essential needs of cartilage regeneration [11]. One of the necessities of transferring the functional properties of native tissue to the product of tissue engineering is the design and production of scaffolds that can mimic the mechanical properties of native tissue. For example, studies have shown that polyethylene glycol and chondroitin sulfate-derived hydrogels produce structures with stiffness gradients (0.005-0.06 MPa) that can mimic the glycosaminoglycan gradient in articular cartilage [19].

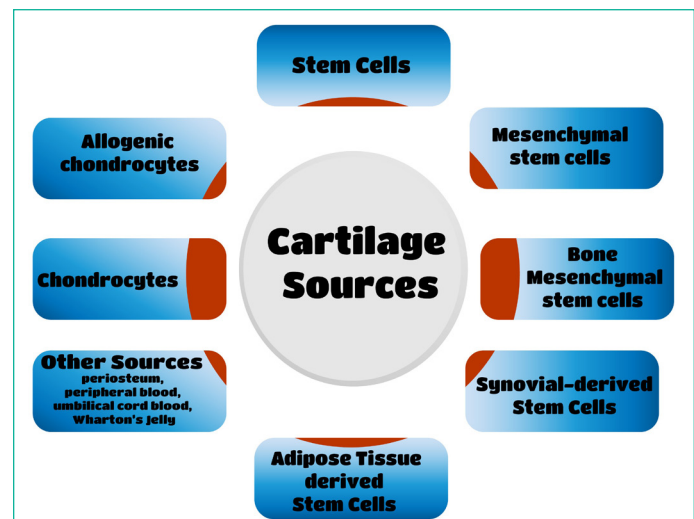


Figure 1: Viable cell sources for cartilage tissue engineering and validated in animal models.

Current Cell Sources for Articular Cartilage Damage

The ultimate goal of cartilage repair is to find an ideal cell source that can be easily isolated, expanded, and cultured to express and synthesize cartilage-specific Extracellular Matrices (ECM), such as type II collagen and aggrecan. Stem cells and chondrocytes have been investigated for their potential as viable cell sources for cartilage tissue engineering and validated in animal models (Figure 1) [20].

Figure 1: viable cell sources for cartilage tissue engineering and validated in animal models

Chondrocytes

Chondrocytes are resident cells of articular cartilage that produce ECM components. As such, they are regarded as the logical choice for cartilage tissue engineering. Indeed, chondrocytes isolated from various tissues, including ribs, nasal septum, ear, or articular cartilage, have been utilized for cartilage tissue engineering [20]. Recently, various studies have focused on articular chondrocytes as a viable cell source for cartilage repair. Autologous Chondrocyte Implantation (ACI) has been regarded as a preferable, long-term treatment prescription. When compared to multipotent marrow cells, passaged articular chondrocytes have the greater innate potential to form hyaline-like cartilage [20]. Nowadays, Matrix-induced Autologous Chondrocyte Implantation (MACI) is used as one of the most extensive methods for the clinical treatment of articular cartilage defects. One major limitation of using chondrocytes is their instability in monolayer culture, which results in their loss of phenotype [21].

As previously mentioned, most efforts to regenerate cartilage have focused primarily on chondrocytes from immature animals. Neonatal and young chondrocytes exhibit faster growth rates, better capacity for rapid expansion *in vitro*, and greater chondrogenic potential compared with chondrocytes isolated from older donors.

However, these limitations can be eliminated by the addition of growth factors such as platelet-derived growth factor (PDGF), Transforming Growth Factor beta 1 (TGF β -1), TGF β -2, fibroblast growth factor 2 (FGF-2), and/or Insulin Growth Factor 1 (IGF-1). Nevertheless, research is still necessary to optimize culture techniques for aged chondrocytes and define their potential clinical uses and limitations [20].

Allogeneic Chondrocytes

Allogeneic chondrocytes have been used with some success in animal models involving rabbits. Although antigens are strongly expressed on chondrocytes, they have been revealed to be non-immunogenic and exert immunomodulatory effects, which make allogeneic chondrocytes an ideal alternative source. The advantages of the allogeneic approach are single surgery, high seeding density with early culture, and decreased dedifferentiated cell use [22]. The isolation of chondrocytes from cartilage ECM in culture causes a loss of the chondrocyte property and results in the conversion of the chondrocytes to fibroblastic cells [23].

Stem Cells

Stem cells are self-renewing cells that can produce more stem cells through mitosis because of their undifferentiated biological character or can differentiate into specialized cells. Stem cells lay a foundation for organ systems and tissues and play various roles in tissue repair, development, and disease progression. They can produce a substantial number of cells, facilitating the restoration of larger defects to help overcome critical challenges in tissue engineering and regenerative medicine [20]. In cartilage regeneration, stem cells are obtained from the patient or other donors and then cultured *in vitro* in a certain microenvironment. Cells are then proliferated and differentiated towards a chondrogenic lineage under a controlled environment, including hypoxia, high-density microenvironment, co-culturing, and specific growth factors. Subsequently, along with bioactive factors and scaffold materials, these cells may be implanted into cartilage defect sites and finally, cartilage regeneration may be achieved [24]. In recent years, the application of MSCs and mesenchymal progenitor cells, including Adipose-Derived Stem Cells (ADSCs), Bone Marrow-Derived Stem Cells (BMSCs), and Embryonic Stem Cells (ESCs) and Induced Pluripotent Stem Cells (iPSCs), has emerged as an attractive strategy to improve the reparative processes of cartilage lesions compared with implantation of other cell types, such as articular chondrocytes. They are regarded as the most suitable or promising cell sources for cartilage tissue engineering and regeneration [20].

Mesenchymal Stem Cells (MSCs)

Mesenchymal Stem Cells (MSCs) are alternative cell sources for cartilage repair. They possess a high proliferation capacity and can be obtained from a variety of tissues, including adipose tissue, bone marrow, periosteum, synovium, umbilical cord vein and placenta [20,23]. MSCs exhibit multilineage potential, such that these cells can differentiate into chondrogenic, osteogenic, and adipogenic lineages *in vitro*. MSCs can undergo chondrogenic differentiation when cultured as micro-pellets in the presence of a defined medium containing TGF β and dexamethasone. Despite MSCs' ability to differentiate into chondrocytes, expansion seems to reduce such ability in these cells. More evidence is needed to observe the efficiency of transplanted MSC-derived chondrocytes in cartilage repair *in vivo*. The transplanted cells induce tissue repair through their secreted soluble factors [23]. MSCs secrete a wide variety of matrix molecules, bioactive factors, growth factors, colony-stimulating factors, cytokines, and chemokines. These secretions can modulate the microenvironment and affect cell migration, proliferation, and differentiation. Furthermore, MSCs are considered "immune privileged" cells, indicating that they may be safer than other cell types for use in cartilage tissue engineering [24]. The pre-clinical data with regards to allogeneic cells is conflicting. Allo-

geneic MSCs in rabbit model have displayed promising potential in comparison with autologous cells. However, autologous chondro-progenitor cells in horse models have been reported to facilitate repair compared with allogeneic cells [25].

Several studies have shown that MSCs are promising cell sources for cartilage regeneration. However, there are some limitations to the use of MSCs. First, the decreasing ability of MSCs for self-renewal with age limits their potential for differentiation. Second, they are associated with a tendency to exhibit tumorigenesis and malignant transformation. Finally, even under controlled conditions, neither the structure nor the function of the restored tissue is similar to that of articular cartilage [24].

BMSCs

With characteristic multipotency, rapid proliferative capacity and ease of isolation, BMSCs are another promising candidate for cartilage regeneration [20]. BMSCs were assessed with respect to cartilage repair in a rabbit model in 1994. In this experiment, a mixture of BMSCs and type I collagen was implanted into the cartilage defects. Twenty-four months later, the histological scores were found to be slightly improved compared with the control group. Since then, increased data from animal models presented optimistic outcomes and confirmed the potential for BMSC applications in cartilage tissue engineering [24]. BMSCs can be differentiated into chondrocytes under different culture conditions. Regardless of culture methods or scaffolds, in terms of *in vitro* culture, TGF β generally enhances chondrogenesis; nevertheless, the degree of chondrogenesis is dependent on the scaffolds [20]. To date, a variety of scaffolds have been used in combination with chondrogenic media supplementation and TGF β to drive chondrogenesis of BMSCs, including agarose, alginate, Polyethylene Glycol (PEG), silk, gelatin/chondroitin/HA tri-copolymer, polyglycolic acid/poly(lactic acid) (PGA/PLA), PLGA-Collagen meshes, electrospun polycaprolactone (PCL), poly(L-lactic acid)-co-poly(ϵ -caprolactone) P(LLACL)/ type I collagen nanofiber yarn mesh, and others [24]. Some reports have shown increased sulfated GAG production by BMSCs cultured in alginate compared with agarose gels [26]. Chondrogenesis was indicated by increased aggrecan and type II collagen accumulation and expression. In addition to TGF β 1, the cycling of GFs (e.g., BMP-6 and IGF-1) also affects chondrogenesis during *in vitro* culture. BMP-2 and TGF β 1 have been proved that they promote the differentiation of MSC into hyaline-like cartilage tissue [24]. BMSC chondrogenesis capacity could be enhanced through hypoxic isolation/expansion. Additionally, the co-culture of BMSCs and chondrocytes is a promising strategy to generate tissue-engineered cartilage yielding increased cell proliferation and cartilaginous ECM deposition with positive type II collagen expression. This may result from cell-cell interactions and GF secretion, or a chondrogenic microenvironment provided by endogenous chondrocytes that increases chondrogenesis of BMSCs [27]. Although comparative trials have shown that BMSC therapy is comparable to ACI and superior to other traditional treatments, Haleem et al. [28] suggested that BMSC application shows better short-to-long-term clinical outcomes as well as fewer complications than ACI. In addition, histological analyses revealed that the defect sites were primarily comprised of hyaline-like cartilage after BMSC implantation. Taken together, these data supported the application of BMSCs in cartilage tissue engineering and regenerative medicine [24]. While BMSCs have been widely used for chondrogenesis, the mechanical integrity of the produced matrix re-

mains a limitation. In long-term MSC-laden agarose gel culture, chondrogenesis was observed, but the mechanical properties and amount of matrix produced were inferior compared with that produced by chondrocytes isolated from the same donor [20]. Animal experiments have utilized allogeneic BMSCs to repair defects in cartilage and bone tissue. Chinese investigators have proved that BMSCs are able to survive, proliferate and differentiate after allo-transplantation, which provides evidence for the application of BMSCs in the tissue engineering for the repair of cartilage defects. Another aim of this study was to prove allogeneic BMSCs potential in clinical settings. Qi et al transfected retroviruses carrying human telomerase reverse transcriptase into BMSCs to prepare the immortalized human BMSCs. Chondrogenic differentiation was induced in the BMSCs in vitro. Therefore, these immortalized BMSCs will be applied in allo-transplantation. The results proved that allogeneic BMSCs survived after allo-transplantation in BMSCs group and differentiated into chondral cells to repair cartilage defects [29].

Unfortunately, the application of BMSCs is limited by several disadvantages. The first is complications involved in harvesting bone marrow from the donor site, including pain, morbidity of the donor site, infection, and even sepsis. Additionally, the limited number of BMSCs and issues related to aging are also factors that should be considered when considering the use of BMSCs in cartilage tissue engineering [30].

Adipose Tissue-Derived Stem Cells (ADSCs)

The adipose tissue is as another source with great promise for multipotent progenitor cells. Similar to the bone marrow, the adipose tissue also originates from the embryonic mesoderm [31]. Adipose tissue is not only easily accessible, also contains a large proportion of MSCs (about 5% of all stromal cells), with a density approximately 100 times more than found in bone marrow [20]. ADSCs, first reported in 2001, exist in all types of white adipose tissues, such as internal and subcutaneous fat, and can be isolated through collagenase digestion [32]. Among the different cell sources, ADSCs have been described an ideal cell source for tissue engineering with the following advantages: the ability to be isolated from tissues in comparatively large quantities, better anti-aging ability in both differentiation and proliferation, excellent anti-inflammatory and non-immunogenic properties, and few ethical concerns in association with application of ADSCs. ADSCs, like BMSCs, can be induced for chondrogenic differentiation in a controlled microenvironment, including a 3D culture environment with a defined medium containing growth factors, such as ascorbic acid, TGF β , and dexamethasone [24]. Recently, ADSC-derived chondrocytes have been achieved in high density micro mass cultures, agarose, alginate, fibrin gel, collagen-based scaffolds, and other substrates [20]. In addition, it has been shown that a novel Elastin Like Polypeptide (ELP) can promote chondrogenic differentiation of ADSCs without media supplements. In monolayer culture, dynamic compression combined with exogenous SRY-related HMG box 9 (SOX9) demonstrated positive effects on chondrogenesis of ADSCs in a 3-D porous poly (lactic-co-glycolic acid) (PLGA) scaffold. The use of GFs, such as Bone Morphogenetic Protein 2 (BMP-2), BMP-4, BMP-6 and FGF-2, also affects the chondrogenesis of ADSCs [33]. Collectively, these results demonstrate the important role of GFs in ADSC-based chondrogenesis. Notably, comparative studies suggest that ADSCs exhibit lower chondrogenic potential compared with stem cells isolated from other sources, such as umbilical cord tissue and bone marrow. As these cells exhibit lower type II collagen

gene expression and reduced accumulation of cartilage-specific matrix proteins compared with other cell types, research is still necessary to optimize the chondrocytic differentiation potential of ADSCs [34]. Furthermore, hypoxia treatment and coculturing can help promote chondrogenic differentiation as well. Previous in vivo studies have reported that applications of ADSCs in cartilage tissue engineering can obtain reliable outcomes. Nonetheless, most of the experiments using ADSCs are from case reports or phase I clinical trials, and only a few control studies have been performed. One multicenter study reported that ADSC injection results in 75% symptom improvement in 63% of patients and about 50% symptom improvement in 91% of patients after 1 year. Nevertheless, additional studies of the use of ADSCs are required to reach definitive conclusions [24].

Synovial-Derived Stem Cells (SDSCs)

The synovial membrane (SM) was recently found to be an attractive source for cartilage tissue engineering owing to its proximity to the articular cartilage. SDSCs were identified as MSCs that could be isolated from the SM while exhibiting multilineage differentiation potential in culture.

Many studies have reported the outstanding potential of SDSCs for chondrogenesis, as well as their applications in cartilage tissue engineering both in vitro and in vivo [35]. By controlling the microenvironment, SDSCs can be induced to form hyaline-like cartilage. Additionally, Koga and coworkers suggested that new cartilage was formed in a rabbit model with transplantation of undifferentiated SDSCs into full-thickness defects of the cartilage. Moreover, ample cartilage matrix secretion was confirmed on the basis of immune-histological scores and transmission electron microscopy. SDSCs and chondrocytes have also been shown to have similar secretion activities. In particular, both may secrete type II collagen without increasing the accumulation of type X collagen and express the gene for proteoglycan [23]. Positive results have also been reported in clinical studies. For example, Sekiya et al. claimed that cartilage regeneration and symptom improvement could be achieved in most patients 3 years after SDSC transplantation through arthroscopy in defected cartilage [35].

Other Sources of MSCs

In addition to above-mentioned tissues, MSCs can also be derived from other connective tissues, such as the skeletal muscle, periosteum, peripheral blood, umbilical cord blood, Wharton's Jelly, and dental pulp. As the largest organ in the human body, skeletal muscle is an attractive cell source for cartilage tissue engineering. Moreover, via muscle biopsy, harvesting of the muscle can be minimally invasive [24]. In 2007, Usas et al. showed that in vivo cartilage healing could be improved by transplanting Muscle-Derived Stem Cells (MDSCs). MSCs can be isolated from the periosteum as well using TGF β , which has the potential to induce the cells towards the chondrogenic lineage. Nevertheless, periosteum-derived MSCs are quite rare and difficult to obtain, contributing to their limited application. Peripheral Blood-Derived Mesenchymal Stem Cells (PBMSCs), which were first isolated in 2000, show comparable chondrogenic potential to BMSCs. Moreover, the quality of cartilage restoration was improved with the use of PBMSC injection in clinical studies [36].

Additionally, extra-embryonic cells, such as Wharton's Jelly Stem Cells (WJSCs), placenta-derived mesenchymal stem cells, and Umbilical Cord Blood Stem Cells (UMSCs) are invaluable

sources for cartilage tissue engineering. For example, Wharton's jelly has been reported as a probable source of chondrogenic potentialized stem cells. The existence of UMSCs was first reported by Erices et al. in 2000; these cells were found to be naiver than BMSCs with greater potential for use in cartilage tissue engineering. Nonetheless, the storage of umbilical cord tissues is challenging, limiting the applications of these cells [37].

Comparison of the Applications of Different MSCs in Cartilage Tissue Engineering

As described above, many MSCs display potential for chondrogenesis; nevertheless, the ideal cell source for MSCs for applications in cartilage tissue engineering is still controversial. Although Bone Marrow-Derived Mesenchymal Stem Cells (BMSCs) are the most widely studied MSCs, synovium-derived MSCs (SDSCs) have superior potential for chondrogenic differentiation. Studies have shown that higher percentages of hyaline cartilage are observed in SDSCs-repaired cartilage tissue [38].

Furthermore, with the use of autologous human serum, SDSCs can be expanded faster than any other cell source. Moreover, pellets derived from the SM show greater secretion of cartilage matrix components, which makes them heavier. ADSCs have better histological appearance and biomechanical properties than periosteum-derived MSCs, but with lower chondrogenic potential than BMSCs. nonetheless, ADSCs are present in adipose tissues at a proportion of 1 in 100 cells, which is approximately 500-fold higher than that of BMSCs in the bone marrow. In contrast, UMSCs show 2 times higher population doubling numbers than BMSCs as well as 1.7 times higher numbers than ADSCs have similar chondrogenic capacity. Unfortunately, the application of UMSCs is limited by difficulties in the storage of umbilical cords [24].

Induced Pluripotent Stem Cells (iPSCs)

As iPSCs possess unlimited self-renewal capacity, they can be used as an infinite cell source for cartilage tissue engineering. The iPSCs, which can be produced from skin fibroblasts by introducing a few key transcription factors, can be used to generate differentiated cells exhibiting young properties, such as healthier production, faster proliferation, and longer-lasting activities. Therefore, they may be a good source of cells to repair articular cartilage defects [20]. Several reports have demonstrated the ability of iPSCs to differentiate into various lineages by overexpression of transgenes or induction with cytokines and small molecules [20,24]. iPSCs can also be combined with 3-D nanofibrous scaffolds for cartilage regeneration. Nam Y et al. had proved that chondrogenic pellets could be generated from the outgrowth cells derived from blood mononuclear cell-derived hiPSCs (CBMC-hiPSCs), and revealed that CBMC-hiPSCs might be regarded as a promising candidate for articular cartilage regeneration. In addition, studies have shown that use of human iPSCs (hiPSCs) can be regarded as a clinically translatable and efficient approach to repair rat osteoarthritic cartilage, with differentiated hiPSCs forming hyaline cartilage tissues 8 weeks after transplantation into the articular cartilage of NOD/SCID mouse knee joints. Chondroinduced hiPSCs were also implanted in osteochondral defects of immunosuppressed rats, showing a great quality of cartilage repair [20]. The preparation of patient-iPSCs and subsequent differentiation under Good Manufacturing Practice (GMP) guidelines is costly. A library of allogeneic clinical GMP grade hiPSCs is under development to lower costs and enable large scale treatment. This iPSC bank is comprised of

cells from homozygous donors for major HLA types in order to minimize immune rejection risk when the generated tissues are transplanted. It is much easier to prepare homozygous HLA hiPSCs than hESCs, because it is easier to find individuals who bear homozygous HLA types and are willing to donate their somatic cells to generate iPSCs in comparison with embryos to generate ESCs. hiPSC - derived chondrocytes tend to resemble chondrocytes of juveniles rather than adults, meaning they possess more anabolic and less antigenic activity. Such findings indicate that a single allogeneic iPSC can be utilized for all recipients, which not only would standardize production quality but prevent excessive costs [23]. Nonetheless, before utilizing iPSCs as a candidate for cartilage tissue engineering, further studies are needed to overcome the drawbacks associated with their use, such as teratoma formation and the possibility of genetic disorders. Nevertheless, due to the tumorigenicity of hiPSCs, their clinical applications are also limited. Although these protocols represent important steps for expanding the use of iPSCs for articular cartilage tissue repair, they still have many limitations. These include use of either feeder or embryoid body stage cells that can abate reproducibility or cause cell heterogeneity. It has been confirmed that the chondrogenically differentiated human iPSCs can form articular cartilage[20].

ESCs

ESCs exhibit the best differentiation potential and may supply an unlimited cell population for cartilage tissue engineering [32]. ESCs can be divided into Wharton's jelly stem cells, umbilical cord blood stem cells, amniotic fluid-derived stem cells and placenta-derived mesenchymal stem cells. Before ESCs differentiate into chondrocytes, these cells must pass through an aggregation stage of embryoid bodies [20]. Transplanted hESCs-derived chondrogenic cells maintain long-term viability with no evidence of tumorigenicity, and provide a highly-efficient, practical and safe strategy of applying hESCs for cartilage tissue engineering. The effects of TGF β and BMP-2 on early stages of chondrogenic differentiation and commitment by hESCs was also evaluated, and found significant chondrogenic induction of hESCs. Mouse ESCs undergo chondrogenesis with BMP-2 and BMP-4, as confirmed by type II collagen and alcian blue staining. Chondrogenesis of ESCs is also upregulated by increased production of endogenous TGF β and BMP signaling activity. Furthermore, co-culture of hESCs with primary chondrocytes was shown to induce chondrogenesis, whereby co-cultured cells expressed type II collagen and SOX9, while cultures of hESCs alone did not. Even though, in clinical, there are many new methods for maintaining hESCs, including serum-free, feeder-free methods, and coculture with other cells. But their applications are extremely limited because of their tumorigenicity [20]. Although the vast proliferation capabilities of ESCs make them an appealing cell source, many limitations continue to hinder their clinical use including difficulties in ESC purity and selection, ethical issues, possibility of teratoma formation and antigenicity [20,24]. Additionally, research using ESCs for cartilage tissue engineering is still relatively new. As such, more information about ESCs, as well as new strategies for differentiation and purification, are required to characterize their potential as a viable cell source for cartilage tissue engineering [39].

Cartilage Progenitor Cells (CPCs)

CPCs are potential alternatives as cell sources for cartilage tissue engineering. Convincing evidence has shown that numerous CPCs can be separated from the surface of the articular cartilage and can be induced to differentiate into cartilage under

a high-density microenvironment using specific growth factors [40]. Moreover, a comparative study indicated that CPCs are superior to MSCs for use in cartilage tissue engineering owing to their lower expression of collagen type X, a sign of hypertrophy of cartilage, however, in cartilage tissue engineering, the major concern associated with the use of CPCs is the difficulty involved in obtaining the cells [24].

Chondrocyte Different Expansion Procedure In Vitro

In the process of articular cartilage tissue engineering, two main phases can be identified. A first phase where few isolated cells need to be expanded in order to provide sufficient cells and a second phase where a cartilage is engineered either inside the body (in vivo) or in a cell culture (in vitro) using an appropriate scaffold. Expanded cells are seeded onto three dimensional scaffolds to form cell-polymer constructs, which are cultured in vitro and then used either as implants or for in vitro research [41].

Retention of phenotypic function in the cell population is critical in cartilage tissue engineering. Articular cartilage derived chondrocytes display minimal proliferative capacity. They dedifferentiate upon repeated passaging and the numbers of cell divisions chondrocytes undergo in vitro decreases with age. The limitations of adult chondrocytes to maintain their phenotype expression and differentiation ability after extensive expansion in vitro has led to the investigation of the potential use of pluripotent stem cells and progenitor cells as a source for tissue engineering [42].

Extensive chondrocyte expansion is required to obtain the number of cells needed for tissue engineering applications. To utilize such cells differentiation would be required to restore their functional phenotype after expansion. Human chondrocytes differentiation has been previously shown to be improved by human serum and growth factors. Because 3 dimensional structures improve phenotype retention in articular chondrocytes as well as encouraging matrix molecules formation, Alginate bead was used for encapsulation as the differentiation method. The differentiation of expanded chondrocytes and chondrogenesis of stem cells is stimulated by the factors including insulin-transferrin-selenium-linoleic acid-bovine serum albumin (ITS + 1) dexamethasone (dex), TGF- β 1, Insulin-like Growth Factor I (IGF-I), and Bone Morphogenic Protein 14 (BMP-14) [43]. Serum may provide chondrogenic factors in a manner dependent on maturation, so that human serum, FBS, and adult bovine serum (ABS) may have different effects on chondrogenesis [44].

Currently Available Culture Systems for Chondrocytes Expansion

Despite the improvements introduced by the use of large scale operation units, monolayer systems which largely rely on simple monolayer culture flasks (i.e., T Flasks, Petri dishes or tissue culture well plates) present very low ratios of surface to volume, which inevitably make them inefficient in term of scalability [45]. If production of articular chondrocytes is to be significantly increased, the number of culture units has to be remarkably increased, making the process time consuming and laborious. The result is that the expansion process may not be cost effective. The introduction of three-dimensional alternative systems including Pellet culture, Encapsulation inside hydrogel beads and microcarriers could potentially provide the improved ratio of surface to volume necessary to cope with the scale of

cell expansion required for allogeneic tissue engineering applications [46]. One of these alternatives is the use of cell-seeded microcarriers for cell expansion. In microcarrier cells culture technology anchorage-dependent animal cells are grown on the surface of small spheres which are maintained in stirred suspension culture. Cells attach and spread on the surface provided by the microcarriers and gradually grow into confluent layers. Finally, the microcarrier expanded cell population has been also proven to maintain the ability to undergo chondrogenesis in vitro, an essential requisite for any proposed expansion method. As a result, due to both its desirable expansion capacity and more critically its operational advantages compared to conventional single layer cultures, micro-carrier cultures are potential alternatives for large scale expansion [47].

Stem Cell Derived ECM for Cartilage Repair

Natural biomaterials such as MSC-ECM have been examined as scaffolds for tissue engineering because of their substantial bioactivity and high biocompatibility [48]. There are new cell-based therapies for cartilage tissue regeneration, including such chondrocyte or stem cell treatment. More than 90% of patients with Autologous Chondrocyte Implantation (ACI) for treating OA had satisfactory function five years after the implantation, according to long-term analysis of clinical trials [49]. In contrast of this, undifferentiated stem cell injections increase the likelihood of cell migration towards the inappropriate cite, and they may develop into ectopic tissue [50]. Increased autoimmune responses may raise the risk of malignancy or other harmful impacts of cell treatment following cartilage regeneration when using modified or allogeneic cells [51]. EVs (Extracellular Vesicles) and soluble substances generated through Mesenchymal Stem Cells (MSC) have a role in stem cell therapeutic efficacy [52]. mRNAs, proteins, microRNAs, and liposomes are all included inside the membrane of endosomal EVs, which have a diameter of 30–150 nm. These characteristics might be useful in the creation of therapeutic biomarkers for medication delivery [53]. As a result of their enhanced physicochemical strength and biocompatibility, EVs such as exosomes have recently emerged as powerful cell-free transfer mechanisms [54]. Apoptosis bodies, Micro Vesicles (MVs), and exosomes are the three primary EV groups released by cells. In addition to miRNA, DNA, RNA, and proteins, several different forms of vesicles can be secreted by MSCs [55]. These vesicle groups are supposed to be homogenous in size and density, although the subtypes are indeed diverse [56]. EVs-MSCs have been shown in preclinical research to be effective in treating a variety of disorders, but clinical trials must still address issues of safety. Human articular chondrocytes or bone marrow stromal cells were used to generate cell type-specific Extracellular Matrix (ECM), which replicates a native milieu, to reduce chondrocyte dedifferentiation during in vitro growth. Spectrometry and atomic microscopy have shown that AC-ECM and BM-ECM have distinct ECM compositions and physical features, respectively. Accumulation of HAC (Human Articular Chondrocytes) cells on AC-ECM were significantly faster than on BM-ECM or the conventional culture surface [57].

Small Molecules (Exosomes)

It has been a long time since Vacanti first introduced the term "tissue engineering" [58], but since then, researchers have used it to develop a new method of regenerating cartilage and bone tissue using a mix of synthetic substrates, cells, and bioactive chemicals with the goal of providing a potential option for improving cartilage and bone regeneration by speeding up the

healing process [59]. In recent years, exosomes, one form of Extracellular Vesicles (EVs) released by cells and that vary in diameter between 50 to 130 nm, have been investigated into it for their wide variety and shedding processes. They were shown to play emerging roles in various cellular mechanisms, such as cell signaling, immune response modulation, and natural cycles of biological colonies [60,61]. Cell-to-cell communication is expected to have a significant impact on the course of healing due to the interconnectedness of chondrocyte regeneration processes. Exosomes, which may deliver bioactive hydrocarbons, nucleic acids, and proteins, are now a hot topic in the regeneration area because of their role in intercellular communication [62]. Coculture of Stem cells with harmed chondrocytes has long been known to promote regeneration through the secretion of numerous factors by SCs, including Proinflammatory cytokines such as IL-6, FGF 2, and insulin-like growth factor, all of which have been shown to promote chondrocyte proliferation and matrix synthesis [63]. In light of the fact that the success of MSC-based joint disease treatments is attributed to paracrine signaling, exosomes, as being one of the components of MSC release, have also been recommended as a successful therapy to restore osteochondral abnormalities [64]. Exosomes from MSCs have been shown to have chondroprotective benefits in joint disorders by Cosenza et al [65]. Chondrocyte indicators (type II collagen and proteoglycans such as aggrecan) and inflammatory markers (such as iNOS) were dramatically increased by intra-articular injection of exosomes from MSCs into a collagenase-induced Osteoarthritis (OA) model. Chondrocyte apoptosis and inflammation worsening by activation of monocytes are two possible outcomes of these substances [66].

Gene therapy: Introducing foreign genetic material or gene patterns into various cell types is known as gene transfer. Gene therapy is use of gene transfer methods to treat illness [67]. Non - viral gene transfer is known as transfection, while viral gene transfer is known as transduction [68]. The initial experiments used ex vivo retroviral gene transduction to modify synoviocytes of patients with terminal rheumatoid arthritis, followed by reinjection of the transformed cells into the Metacarpophalangeal (MCP) joint [69,70]. Concerning OA, a phase I procedure is presently underway, employing an ex vivo method utilizing retroviral TGF- β [71]. There is a growing amount of evidence from a variety of transgenic and somatic genetic manipulation research suggesting gene transfer may be used to increase athletic capacity. Numerous genes have previously been cloned into functional vectors, and others are being examined for clinical trials in order to disease therapy [72]. The difficulty for antidoping authorities will be detecting these endogenously created gene products because to the similarities between the transferred cDNA and the endogenously manufactured protein, as well as the low specificity of indirect testing technologies [73]. The reports have concentrated on membrane healing proteins, transposable elements, and growth factors delivery. Much of the research has been on IGF1 expression. Regardless of the vector utilized, the primary limiting factor in the efficiency of cartilage regeneration following GT has been temporary transcription of the gene product. A second reason limiting GT's effects in cartilage healing has been the achievement of inadequate amounts of target proteins such as growth pathway. But at the other hand, in the initial few days following GT, supra-therapeutic or hazardous doses are typically reported in an attempt to boost the target protein's expression. Despite the advances, the length of expression is still inadequate to induce cartilage regeneration [74].

Transplantation Methods in Cartilage Tissue Engineering

Tissue engineering involving cells seeded on scaffolds made of synthetic and natural biodegradable polymers can be a promising for the future treatment of diseases including cartilage defects. These polymers can be injected as a minimally invasive procedure or as a pre-made graft to treat large irreparable defect, including osteoarthritis. Scaffolding as a mechanical substrate for cells and bioactive factors could help guide and organize cells to regeneration process [75]. Today, various surgical and non-surgical methods are used to cartilage regeneration. In any method of cartilage regeneration, preparation of a suitable substrate for replacing the implants is very important [2]. Several new surgical procedures have been developed to promote cartilage regeneration.

Injection method: Minimally invasive surgery can reduce connective tissue damage and wound size, resulting in faster healing, so we have tried to design injectable structures that replace surgical techniques to repair cartilage tissue [76]. There are several cross linking manner for in situ polymerization of injectable structures, such as thermal, chemical and photo-cross linking [77]. Photo-cross linker has more considered than other polymerization methods due to possibility of spatial and temporal control of the polymerization method by adjusting the light intensity and duration of light exposure [78]. These cross-linking methods have also been used for cell encapsulation and synthesis of bioactive molecules [79,80]. In addition to being less invasive, these structures may fill gaps, especially for the treatment of irregular form or difficult to reach defects [80].

Hydrogels are common and injectable scaffolds in cartilage tissue engineering [81]. The production of new tissue with this method is enhanced by implanting cells in a 3-D matrix. Hyaluronic Acid (HA) and collagen based materials are widely used as biodegradable and biocompatible material for cartilage regeneration [82]. In addition, several natural, synthetic materials and copolymers have been shown to mimic the mechanical environment of cartilage tissue and its properties [83]. In recent study, the role of advanced manufactured methods in cartilage regeneration was evaluated in cartilage defects of rabbit model. They used methacrylate hyaluronic acid as a photo cross linker hydrogel containing Kartogen in with PLGA nanoparticle. The results of this study showed that in comparison with the untreated group, this single-step surgery without cells, after 3 months was able to show the formation of hyaline cartilage with a high content of type II collagen [84]. Microspheres have been considered as an efficient transport system for the controlled release of drugs and biological agents. In tissue engineering, microspheres are widely used as cell-carrying scaffolds [85]. Currently, intra-articular injections of microspheres are used as a minimally invasive method in the treatment of cartilage defects. Cytodex 1, CultiSpher S and SphereCol are some samples of commercial microspheres that are used for cartilage regeneration [86,87]. Intra-articular injection of microspheres containing growth factors to treat cartilage defects like osteoarthritis is another application of microspheres used in the clinic. Platelet-Rich Plasma (PRP), Fibroblast Growth Factor (FGF), and Transforming Growth Factor-1 (TGF-1), are some biological factors that were injected to stimulate proliferation of chondrocyte or chondrogenic differentiation from stem cells. As well as anti-inflammatory cytokines, are used to repress arthritis-related inflammation [87].

Arthroscopic method: Arthroscopic surgery is used as an orthopedic surgery method to diagnose and treatment of articu-

lar defects. In this procedure, the size and shape of the cartilage defect are measured using an arthroscopically graded probe [88]. In addition, many operations can be performed by arthroscopy. It's appropriate for moderate cartilage defects. Among the advantages of this method is the low cost of arthroscopy operation and minimizing the invasion and thus improving rehabilitation. In previous study, by arthroscopic surgery method, several cartilage matrices were isolated from two cadaver cartilage and used in a hip model. Cartilage implants based on cell-free Polyglycolic Acid- (PGA-) hyaluronan scaffold with a flat surface were implanted by with a thickness of 10 mm × 15 mm. The implants were fixed on the hip cartilage defect by fibrin glue. The results have been shown the implant was reinforced by PGA scaffolds [89].

Open surgery method: Open surgery as a common method widely applied in cartilage tissue engineering. Although arthroscopy is less invasive, but in some cases, such as cartilage defects in areas like the posterior femoral condyle, patella and larynx, as well as arthroscopic surgery for some scaffolds is not applicable [88]. Scaffolds designed by tissue engineering are typically transplanted through open surgery. These scaffolds are made by various techniques such as foam casting, electrospinning [90], solvent casting and particle leaching [91], faze separation as well as 3D bioprinting before operating [92]. 3D bioprinting implants contain a wide range of materials and compounds such as calcium polyphosphate and PVA, hydroxyapatite and calcium phosphate, tricalcium phosphate, PCL and chitosan, collagen, and mesenchymal cells are grafted by open surgery for cartilage regeneration [80]. Biopen is a novel 3D bioprinting machine that allows surgery to print bionic and cells during surgery, this technique can revolution in cartilage regeneration. Simultaneous in situ printing of scaffolds and cells on cartilage defect, performed in an open surgical session. Bella et al. [93] used biopen for cartilage regeneration un sheep model. HA-GelMaBioink, was used as a shell layer and allogeneic fat-derived mesenchymal stem cells as a core layer. The material and cells simultaneously were printed on the lateral and internal condyles of both femurs. The results indicated that in situ printing of cell and scaffold simultaneously on cartilage defect had no side effects, in all cases. Biopen can be used in a similar way like other surgical instruments. The shape and thickness of the 3D printed material as well as the speed of printing can be control by surgeon. Biopen could produce a 3D-printed scaffold that completely matched the profundity and shape of defects.

Microfracture surgery method: Microfracture method as a surgical procedure is used to treat cartilage defects. In this method, by making several small holes in the bone under the cartilage defect, the Mesenchymal Stem Cells (MSCs) are stimulated and oriented from the bone marrow and differentiate into cartilage cells [94]. Steadman et al. [94] was designed this method in the early 1980s. At present, micro-fracture surgery is considered as an alternative treatment instead of open surgery for full-thickness cartilage disorder [95]. According to the research, cartilage regeneration was done better in young than older patients [96]. This method can also be used to transplant clinical engineering products. In a study to improve the regeneration process of cartilage repair, 3 D printed scaffolds from aggrecan without cells were grafted with micro-fracture method in a Lapin model [97,98]. The results showed that aggrecan has a high potential for cartilage regeneration and this surgical method was effective and cost effective [99].

Mosaicplasty method: Mosaicplasty is another method for

cartilage regeneration. Hangody and el al. [100] was designed this method in 1992 for clinical usage. This surgery method is the mosaic like implantation of some tiny cylindrical plugs of osteochondral to regenerate a cartilage surface [101]. Cartilage grafting from a place that is not under mechanical pressure to place that is exposed to mechanical load is prone to damage due to structural and mechanical differences between the two areas of cartilage [102]. Osteochondral allograft can be resolve this problem, but the recipient's immune reaction remained [103]. Bartha et al. [104] evaluates porous Poly (Ethylene Oxide) Terephthalate / Poly (Butylene Terephthalate) (PEOT / PBT) implants to repair the cartilage defect. PEOT / PBT implants were successfully transplanted and reduced donor complications, bleeding, and inflammation after mosaicplasty surgery.

Available Products

Business research predicts that the market for cartilage repair products will grow definitely between 2020 and 2027. Data Bridge Market Research analyses the market to account for USD 4.5 billion by 2027 growing at a CAGR of 6.30% in the above-mentioned forecast period. Increasing sports injuries, obesity, a growth in the elderly population and raise awareness of osteoarthritis will be important reasons for this development in market size. Of course, the high cost of repair procedures, inappropriate surveillance policies, and problems with the insurance and repayment system will be obstacles to market growth. Cartilage repair is known as treatments used for damaged cartilage which can cause pain and inability to move for patients. The main joints involved are the hip, knee, ankle and spine. Analgesics and some medications are used for this purpose but do not produce a significant therapeutic response. The cartilage repair market is segmented on the basis of the type of cartilage, treatment modality, treatment type, application site, and surgical procedure. These segments will help you analyze valuable market overview and market insights to help you in making strategic decisions for the identification of core market applications.

- On the basis type of cartilage, the cartilage repair market is divided into fibrocartilage, hyaline cartilage and other
- Based on treatment modality, the cartilage repair market is divided into cell- based and non- cell based. Cell based is divided into smaller subgroups: chondrocyte transplantation and growth factor technology. The non-cell-based segment consists of tissue scaffolds and cell- free composites.
- The treatment type part of the cartilage repair market is divided into palliative and intrinsic repair stimulus. Palliative segment itself includes 2 sections: viscosupplementation and debridement & lavage
- Application segment of the cartilage repair market includes different parts of the body namely knee, spine, ankle, hip and others
- Based on surgical procedure, the cartilage repair market is segmented into micro fracture, debridement, abrasion arthroplasty, autologous chondrocyte implantation, osteochondral autograft and allograft transplantation, allogeneic chondrocyte implantation, cell-based cartilage resurfacing and others

the U.S., Canada, and Mexico in North America, Germany, France, U.K., Netherlands, Switzerland, Belgium, Russia, Italy, Spain, Turkey, Rest of Europe in Europe, China, Japan, India, South Korea, Singapore, Malaysia, Australia, Thailand, Indone

Table 1: Cartilage products available in the market.

| Company | Product | Description | References | Country |
|-------------------------------------|---|---|------------------------------|------------------------------|
| Anika Therapeutics SRL | HYALOFAST® | HYALOFAST non-woven 2×2 cm or 5×5 cm biodegradable hyaluronic acid-based scaffold. Single 3D fibrous layer of HYAFF®, a benzyl ester of hyaluronic acid (HA), mesenchymal stem cells (MSCs), embryonic-like environment, Bio-resorbable and strong safety profile | [105-107] | USA SRL Italy |
| Osiris Therapeutics, Inc. | Cartiform® | Cartiform®: cryopreserved viable osteochondral allograft, 3-dimensional scaffold of hyaline cartilage. Improve the healing potential of bone marrow stimulation procedures. Mesenchymal stem cell (MSC). | [108, 109] | USA |
| Arthrex, Inc. | OATS® (As part of the Allograft) | Dovetail Meniscal Allograft Set/ The technique used with the Dovetail Meniscal Transplant Set creates a trapezoidal bone block allograft. | [110-112] | Florida/ USA |
| BioTissue | BioSeed®-C Chondrotissue® | BioSeed®-C autologous 3-dimensional graft for chondrocyte with maximum mechanical resistance and form stability, compare to other collagen or gel-like grafts Chondrotissue® a one-step, CE marked, cell-free implant Stem cells are obtained by performing standard marrow stimulating procedures, such as micro fracturing or Priddle drilling. | [113] [114] | Germany |
| Lifenet health | FlexiGRAFT® meniscus Matrigraft® | Meniscus transplantation/ Processed with tibial bone block to allow for surgical technique flexibility such as double bone plug technique, keyhole technique or dove tail technique. Consists of two types of shafts and fibular wedge. Fibular wedge type is Natural shaped, parallel fibular wedge, Shafts Matrigraft consists of Cortical/cancellous shafts, designed to provide immediate structural support to restore segmental bone loss. Joint Arthroplasty, Tumor Resection and Reconstruction, Fracture Management, Deformity Correction, Corpectomy, Anterior Cervical Fusion | [115] [116] | Virginia/ USA |
| B. Braun Melsungen AG | Novocart® Basic | Novocart® Basic biphasic, three-dimensional collagen-based matrix consists of a collagen membrane cover and a collagen sponge lying underneath. Biomaterials of bovine origin. Chondral and osteochondral lesions of grade III-IV° (ICRS Classification) – for smaller defects Focal, traumatic defects and Osteochondrosis dissecans | [117] | Germany |
| ConMed Corporation | CartiMax Osteochondral Allografts Cartilage Allograft Matrix | CartiMax® is viable, cartilage fibers combined with cartilage allograft matrix to make a biologically-active scaffold with putty-like handling characteristics used to treat focal cartilage defects. Available exclusively from CONMED through MTF Biologics, MOPSTM (Missouri Osteochondral Preservation System) preservation and storage services create osteochondral allografts with consistently high viable chondrocyte density. Taken from the distal femur and processed using MTF's proprietary methods | [118] [119, 120] [121] | NY/ USA |
| Swedish Orphan Biovitrum AB SOBI | ChondroCelect | Chondro celect the first cell-based product to be approved in Europe which is a medicinal product for use in autologous chondrocyte. This product has been approved by the European Medicines Agency with Agency product number EMEA/H/C/000878 | [122] | Swedish Belgium |
| Medipost | CARTISTEM® | Allogeneic umbilical cord blood-derived mesenchymal stem cells has been market-approved with Biologics License Application (BLA) for commercial sale by the Ministry of Food & Drug Safety (MFDS) in January 2012. | [123] | Korea |
| Histogenics | Neocart | 1NeoCart harvesting cartilage cells from the non-weight-bearing cartilage surface of the patient's femur. Histogen research for Neocart trial seems to have failed and stopped to reach endpoints[1] | [124] | USA |
| MEDTRONIC | | Infuse™ Bone Graft, Grafton™ demineralized bone matrix (DBM), Magnifuse™ Bone Graft, The Master-graft™ are some examples for their products. | | Ireland |
| Stryker | Prochondrix CR | ProChondrixCR cryopreserved, fresh osteochondral allograft contains live cells and biological components | 1- [125] | USA |
| Smith & Nephew PLC | CARGEL | CARGEL Bio-scaffold single-step bone marrow stimulation procedure. mixing a buffer, a chitosan solution and the patient's whole blood | [126, 127] | UK |
| Vericel Corporation | MACI®(autologous cultured chondrocytes on porcine collagen membrane) | The symptomatic repair, full-thickness cartilage defects of the knee in adult patients. FDA approved | [128] | Europe USA |
| Zimmer Biomet Holdings, Inc | Chondrofix (Osteochondral Allograft) DeNovo NT | Minimally manipulated human tissue graft shelf-stable graft. Immediate post-operative weight bearing, DeNovo®NT Natural Tissue Graft off-the-shelf human tissue, reducing the need for periosteal flap unlike ACI | [129] [130] | Switzerland |
| Orthox | FibroFix™ Cartilage | | [131] | UK |
| Allosource | ProChondrix® CR | ProChondrix CR, a laser-etched, fresh cryopreserved osteochondral allograft Presence of native growth factors and Viable chondrocytes | [125] | CO/ USA |
| Cartiheal | Agili-C | Agili-C™ is a cell-free, off-the-shelf implant porous, biocompatible, and resorbable bi-phasic scaffold, | [132] | Israel New Jersey/ USA |
| MTF Biologics | Profile® Cartimax | MTF Biologics' line of costal human cartilage grafts are primarily used for revision rhinoplasty procedures, both reconstructive (post-trauma or Mohs procedure for basal cell and squamous cell carcinoma of the nose), and cosmetic. Grafts are available in both segment and thinner, pre-cut sheet (Profile®) forms. CartiMax is a ready-to-use, off-the-shelf viable cartilage allograft that can fill cartilage defects up to a 5cm ² lesion. MTF Biologics, Research and Development Department. | [133] [134] | USA |
| Collagen Solutions PLC | Collagen scaffolds | Collagen scaffolds are ideal for repairing hyaline joints due to their perforated structure and reconstructive structure. | [135, 136] | London/UK |

| | | | | |
|--------------------------|---|--|----------------|-----------------|
| RTI Surgical, Inc. | Fresh-Stored Osteochondral Grafts Allograft Dermis | Implants includes Fresh-stored OC Femoral Condyle, Fresh-stored OC Talus, Fresh-stored OC Humeral Head, Fresh-stored OC Femoral Distal Tibia, Fresh-stored OC Femoral Trochlea and Fresh-stored OC Femoral Patella Matrix HD® Allograft Dermis is an acellular human dermis allograft sterilized to a Sterility Assurance Level (SAL) of 10 ⁻⁶ using the Tutoplast® Tissue Sterilization Process. The three-dimensional intertwined multidirectional fibers and mechanical properties of the native dermis tissue. | [137] [138] | USA |
| JRF ortho | osteocondral allografts | Femoral Condyle, Precut Fresh OCA Cores (great alternative for focal lesions of up to 20 mm or for Autograft OATS® backfill.), Bi-Compartment Allograft (Combination Femoral Trochlea and Condyle), Custom Allografts (joint, and size-specific osteochondral grafts), Distal Tibia(for resurfacing ankle or shoulder), Femoral Head, Femoral Trochlea (utilized for reconstructing the complex curvature of the trochlear region), Humeral Head(shoulder), Metatarsal Bone (toe), Patella Bone, Talus, Tibia Plateau (with attached Meniscus), Whole Femoral Condyle (allograft includes the entire distal femur) and Whole Tibia Plateau (with intact Menisci) | [139] | USA Colorado |
| NANOCHON | 3D print-nano implant | Using for treat tears, sports injuries, early onset osteoarthritis and other forms of full-thickness cartilage loss. 3D printed from a novel, nanostructured synthetic material. Recently Nanochon was awarded a Phase I SBIR from the National Science Foundation, and is planning to undertake a human study in 2019. | [140] | US Study |
| Regentis biomaterials | Gelrin C hydrogel | After standard microfracture, the hydrogel is injected as a liquid, conforming to the lesion size, shape and depth. Hydrogel Implant GelrinC Demonstrates Impressive Recovery Rates for Patients with Knee Cartilage Damage. Besides there is an ongoing clinical trial about this product. | [141] | Israel |
| Cytex | 3-Dimensional woven implants | Bone marrow derived mesenchymal cells The implant includes a three-dimensional woven textile scaffold and a three-dimensional rigid, porous substrate. | [142] | US |
| DePuy Synthes | COR® Autograft | The Arthroscopic Technique for Repair of Osteochondral Defects Which focuses on the extraction of graft tissue in the autograft process | [143] | Indiana USA |
| Azellon cell therapeutic | Azellon's (Stem cell on membrane) | Azellon's meniscal repair technology will combine patient's bone marrow stem cells with a special membrane that helps to deliver the cells into the injured site. | [144] | UK |
| Educell | ChondroArt TM | ChondroArt TM are tissue-engineered products for cartilage repair in knee and other joints, based on implantation of autologous chondrocytes. | [145] | Slovenia |

sia, Philippines, Rest of Asia-Pacific (APAC) in the Asia-Pacific (APAC), Saudi Arabia, U.A.E, South Africa, Egypt, Israel, Rest of the Middle East and Africa (MEA) as a part of the Middle East and Africa (MEA), Brazil, Argentina and Rest of South America as part of South America are countries covered in cartilage market reporting. North America is expected to overtake the cartilage engineering market between 2020 and 2027 (adapted from <https://www.databridgemarketresearch.com>)

In this section, we decide to describe some trend products available in the market in the field of cartilage allogeneic transplantation and biological scaffolds for chondrocytes.

At the end, we introduce some famous company with autologous products. As shown at mccourier.com and our searches, the major players covered in the cartilage repair market report are Histogenics, Vericel, Swedish Orphan Biovitrum AB., MEDIPOST, Zimmer Biomet, Osiris, B. Braun Melsungen AG., Stryker, Smith & Nephew, Medtronic, CONMED Corporation., Arthrex, Life Net Health, Anika Therapeutics, Inc., BioTissue, among other domestic and global players. At the end, we report on several other companies and products in table 1. (Adapted from <https://www.mccourier.com>).

Table 1: Cartilage products available in the market

Rejection of allograft transplantation due to the immune response remains a common and serious challenge in allogeneic tissue or organ transplantation that leads to loss of durability [146]. In tissue engineering, MSCs used on 3D scaffolds can adjust the immune system by reducing inflammatory agents and increasing the release of anti-inflammatory cytokines like Prostaglandin E2 (PGE2), Transforming Growth Factor- β (TGF- β), and Hepatocyte Growth Factor (HGF) [147]. Increase amount of TGF- β and other anti-inflammatory cytokines could induced the regulatory T cells (Tregs) [148] and macrophages activity

[149]. MSCs with immunosuppressive properties can directly inhibit the activity of immune cells. The binding of Fas Ligand (Fas-L) and Programmed Death-Ligand 1 (PD-L1) on the surface of MSCs to surface receptors of immune cells, lead to decrease the immune responses [150].

The selected 3D scaffolds in tissue engineering, in addition of mimic the mechanical and physical properties of the tissue, should also not stimulate inflammatory agents [146]. Natural biomaterials like collagen have more biocompatibility and minor immunogenicity [146,151]. Some natural biomaterials are inherently anti-inflammatory, including high molecular weight Hyaluronic Acid (HA) and chitosan [152], which can decrease the type of reactive oxygen [153]. However, the use of anti-inflammatory drugs is common for most biomaterials [154]. According to studies, the structure, shape and geometry of the scaffold can also affect inflammation. In an experimental study it was shown that spherical material was implanted in various biological materials have better biocompatibility and the amount of fibrosis and FBR depend on the dimension of the material [155,156]. In addition, the researches have been shown that MSCs enclosed in 3D scaffolds due to the microstructure of scaffold had less inflammation compared to Two-Dimensional (2D) culture after transplantation. In three-dimensional culture, the amount of macrophages was reduced and production of anti-inflammatory proteins like PGE2 and TSG-6 were increased [156].

Yang et al. [157] to overcome immune response challenge in osteochondral grafting, used basal Fibroblast Growth Factor (bFGF) in combination with agarose gel to modulate and control full-thickness cartilage grafting. This procedure could reduce inflammation at the graft site and prolonged the survival of allogeneic cartilage implants [157]. This study has shown that bFGF can prevent the activation of inflammatory factors that lead to

the secretion of cytokines and the destruction of transplanted tissue. Also bFGF increased the levels of CD4+, CD25+, Foxp3+ and regulatory T cells (Tregs) in the recipient blood. Increasing Tregs can protect implanted tissues against immune rejection [158].

In another method using immunosuppressive drugs simultaneous with allograft Mesenchymal Stem Cell (MSC) transplantation can reduce the host immune response. Immunosuppressive drugs could modify the mesenchymal stem cells efficacy by extending the viability of allograft tissue or organ transplants, and in turn, mesenchymal cells can control the side effects of immunosuppressive drugs [159,160]. Ge et al. [160] used the Rapa to suppress the immune responses of allograft- MSC transplantation. Previous studies have shown that allograft-MSC genetic modification can be effective in controlling inflammation [161]. The MHC-1 expression decreased by US11 gene modification in bone marrow derived MSC. This modification also led to the protection of MSC by cytotoxic lymphocytes and prolonged the survival of mesenchymal stem cells in the allogeneic receptor [162]. Chen et al. [163] showed that genetic modification of mesenchymal stem cells could control the inflammation of osteoarthritis after allograft transplantation. Previous studies have shown that articular inflammation activates the IL-1b and TNF- α pathways and inhibits the chondrogenesis of mesenchymal stem cells [164], and can prevent fusion of grafted cartilage tissue [165]. Based on studies to control inflammation after allograft transplantation, use of IL-1b inhibitory growth factors such as IGF-1, Platelet-Derived Growth Factor (PDGF) -bb, Bone Morphogenetic Proteins (BMP-2 and BMP-9), GAG compounds (such as glucosamine), hyaluronic acid, and chondroitin sulfate, as well as Platelet-Rich Plasma (PRP), have been shown to be effective [166,167].

The use of PRP as an autologous source containing chemokines / cytokines, adhesive proteins, and effective growth factors for tissue repair is increasing in the therapeutic system. By inducing the synthesis of hyaluronic acid, proteoglycans and collagen type II have a protective effect on chondrocytes and stimulates their proliferation [168,169]. It also has anti-inflammatory and immunosuppressive effects by inhibiting macrophages as well as suppressing inflammatory factors such as metalloproteinases [170,171].

Conclusions

In summary, reconstruction of articular cartilage defects is a complex procedure. Despite the efforts of researchers, there is still no effective and long-term treatment for articular cartilage damage. Common treatments include surgical procedures such as debridement and arthroscopy, chondrocyte implantation, plastic mosaic, micro fracture, periosteal transplant, heart transplant, osteotomy, and bone marrow stimulation. Cartilage tissue engineering approaches by using different stem cell sources along with appropriate biological scaffolds, chondrogenic factors and physical stimuli, can be a promising way to overcome current limitations and cartilage reconstruction.

Researchers are trying to advance current cartilage therapies toward a consistently successful approach for articular cartilage regenerating. Despite many advances, tissue engineering techniques have limitations for clinical applications, the main problem being in terms of translation, modulation of the host immune system, transplant behavior in the host body, and recovery steps.

Genetic engineering, 3D bioprinting method and cell therapy are being developed alongside other technologies. In the future, combining current strategies with tissue engineering approaches could be a viable solution for the final treatment of cartilage defects.

Author Statements

Author Contributions

All authors have contributed equally to the article conception, data gathering, preparation, draft writing, image drawing, and submission processes.

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Declarations of Interest

The authors have no financial conflicts of interest.

References

1. Chu CR, Convery FR, Akeson WH, Meyers M, Amiel D. Articular cartilage transplantation: clinical results in the knee. *Clinical orthopaedics and related Research*. 1999; 360: 159-68.
2. Haleem AM, Chu CR. Advances in tissue engineering techniques for articular cartilage repair. *Oper Tech Orthop*. 2010; 20: 76-89.
3. Banani MA, Rahmatullah M, Farhan N, Hancox Z, Yousaf S, Arabpour Z, et al. Adipose tissue-derived mesenchymal stem cells for breast tissue regeneration. *Regen Med*. 2021; 16: 47-70.
4. Arabpour Z, et al. Designing biomaterials for regenerative medicine: state-of-the-Art and future Perspectives. 2020.
5. Fathi A, Khanmohammadi M, Goodarzi A, Foroutani L, Mobarakkeh ZT, Saremi J, et al. Fabrication of chitosan-polyvinyl alcohol and silk electrospun fiber seeded with differentiated keratinocyte for skin tissue regeneration in animal wound model. *J Biol Eng*. 2020; 14: 27.
6. Liu Y, Zhou G, Cao Y. Recent progress in cartilage tissue engineering—our experience and future directions. *Engineering*. 2017; 3: 28-35.
7. Bhattacharjee M, Coburn J, Centola M, Murab S, Barbero A, Kaplan DL, et al. Tissue engineering strategies to study cartilage development, degeneration and regeneration. *Adv Drug Deliv Rev*. 2015; 84: 107-22.
8. Rowland CR, Colucci LA, Guilak F. Fabrication of anatomically shaped cartilage constructs using decellularized cartilage-derived matrix scaffolds. *Biomaterials*. 2016; 91: 57-72.
9. Kwon H, Brown WE, Lee CA, Wang D, Paschos N, Hu JC, et al. Surgical and tissue engineering strategies for articular cartilage and meniscus repair. *Nat Rev Rheumatol*. 2019; 15: 550-70.
10. Shen S, Chen M, Guo W, Li H, Li X, Huang S, et al. Three dimensional printing-based strategies for functional cartilage regeneration. *Tissue Eng Part B Rev*. 2019; 25: 187-201.
11. Eslahi N, Abdorahim M, Simchi A. Smart polymeric hydrogels for cartilage tissue engineering: a review on the chemistry and biological functions. *Biomacromolecules*. 2016; 17: 3441-63.
12. Techawanitchai P, Ebara M, Idota N, Asoh T, Kikuchi A, Aoyagi T. Photo-switchable control of pH-responsive actuators via pH jump reaction. *Soft Matter*. 2012; 8: 2844-51.
13. Klouda L, Mikos AG. Thermoresponsive hydrogels in biomedical applications. *Eur J Pharm Biopharm*. 2008; 68: 34-45.

14. Cho JK, Hong KY, Park JW, Yang HK, Song SC. Injectable delivery system of 2-methoxyestradiol for breast cancer therapy using biodegradable thermosensitive poly (organophosphazene) hydrogel. *J Drug Target*. 2011; 19: 270-80.
15. Naficy S, Spinks GM, Wallace GG. Thin, tough, pH-sensitive hydrogel films with rapid load recovery. *ACS Appl Mater Interfaces*. 2014; 6: 4109-14.
16. Schulz DN, Peiffer DG, Agarwal PK, Larabee J, Kaladas JJ, Soni L, et al. Phase behaviour and solution properties of sulphobetaine polymers. *Polymer*. 1986; 27: 1734-42.
17. Tomatsu I, Peng K, Kros A. Photoresponsive hydrogels for biomedical applications. *Adv Drug Deliv Rev*. 2011; 63: 1257-66.
18. Yoshida R, Sakai K, Okano T, Sakurai Y. Pulsatile drug delivery systems using hydrogels. *Adv Drug Deliv Rev*. 1993; 11: 85-108.
19. Zhu D, Tong X, Trinh P, Yang F. Mimicking cartilage tissue zonal organization by engineering tissue-scale gradient hydrogels as 3D cell niche. *Tissue Eng Part A*. 2018; 24: 1-10.
20. Huang K, Li Q, Li Y, Yao Z, Luo D, Rao P, et al. Cartilage tissue regeneration: the roles of cells, stimulating factors and scaffolds. *Curr Stem Cell Res Ther*. 2018; 13: 547-67.
21. Erickson BJ, Strickland SM, Gomoll AH. Indications, techniques, outcomes for matrix-induced autologous chondrocyte implantation (MACI). *Oper Tech Sports Med*. 2018; 26: 175-82.
22. Boopalan PRJVC, Varghese VD, Sathishkumar S, Arumugam S, Amarnath V. Similar regeneration of articular cartilage defects with autologous & allogenic chondrocytes in a rabbit model. *Indian J Med Res*. 2019; 149: 650-5.
23. Yamashita A, Tamamura Y, Morioka M, Karagiannis P, Shima N, Tsumaki N. Considerations in hiPSC-derived cartilage for articular cartilage repair. *Inflamm Regen*. 2018; 38: 17.
24. Ma Q, Liao J, Cai X. Different sources of stem cells and their application in cartilage tissue engineering. *Curr Stem Cell Res Ther*. 2018; 13: 568-75.
25. Goldberg A, Mitchell K, Soans J, Kim L, Zaidi R. The use of mesenchymal stem cells for cartilage repair and regeneration: a systematic review. *J Orthop Surg Res*. 2017; 12: 39.
26. Connelly JT, García AJ, Levenston ME. Inhibition of in vitro chondrogenesis in RGD-modified three-dimensional alginate gels. *Biomaterials*. 2007; 28: 1071-83.
27. Qing C, Wei-ding C, Wei-min F. Co-culture of chondrocytes and bone marrow mesenchymal stem cells in vitro enhances the expression of cartilaginous extracellular matrix components. *Braz J Med Biol Res*. 2011; 44: 303-10.
28. Migliorini F, Berton A, Salvatore G, Candela V, Khan W, Longo UG, et al. Autologous chondrocyte implantation and mesenchymal stem cells for the treatments of chondral defects of the knee-A systematic review. *Curr Stem Cell Res Ther*. 2020; 15: 547-56.
29. Tu Q, Valverde P, Li S, Zhang J, Yang P, Chen J. Osterix overexpression in mesenchymal stem cells stimulates healing of critical-sized defects in murine calvarial bone. *Tissue Eng*. 2007; 13: 2431-40.
30. Nie C, Yang D, Morris SF. Local delivery of adipose-derived stem cells via acellular dermal matrix as a scaffold: a new promising strategy to accelerate wound healing. *Med Hypotheses*. 2009; 72: 679-82.
31. Jezierska-Woźniak K, Nosarzewska D, Tutas A, Mikołajczyk A, Okliński M, Jurkowski MK. Use of adipose tissue as a source of mesenchymal stem cells. *Postepy Hig Med Dosw (Online)*. 2010; 64: 326-32.
32. Ma Q, Liao J, Cai X. Different sources of stem cells and their application in cartilage tissue engineering. *Curr Stem Cell Res Ther*. 2018; 13: 568-75.
33. Huang K, Li Q, Li Y, Yao Z, Luo D, Rao P, et al. Cartilage tissue regeneration: the roles of cells, stimulating factors and scaffolds. *Curr Stem Cell Res Ther*. 2018; 13: 547-67.
34. Chen JL, Duan L, Zhu W, Xiong J, Wang D. Extracellular matrix production in vitro in cartilage tissue engineering. *J Transl Med*. 2014; 12: 88.
35. Sekiya I, Muneta T, Horie M, Koga H. Arthroscopic transplantation of synovial stem cells improves clinical outcomes in knees with cartilage defects. *Clin Orthop Relat Res*. 2015; 473: 2316-26.
36. Usas A, Huard J. Muscle-derived stem cells for tissue engineering and regenerative therapy. *Biomaterials*. 2007; 28: 5401-6.
37. Erices A, Conget P, Minguell JJ. Mesenchymal progenitor cells in human umbilical cord blood. *Br J Haematol*. 2000; 109: 235-42.
38. Cui D, Li H, Xu X, Ye L, Zhou X, Zheng L, et al. Mesenchymal stem cells for cartilage regeneration of TMJ osteoarthritis. *Stem Cells Int*. 2017; 2017: 5979741.
39. Toh WS, Lee EH, Cao T. Potential of human embryonic stem cells in cartilage tissue engineering and regenerative medicine. *Stem Cell Rev Rep*. 2011; 7: 544-59.
40. Wang R, Jiang W, Zhang L, Xie S, Zhang S, Yuan S, et al. Intra-articular delivery of extracellular vesicles secreted by chondrogenic progenitor cells from MRL/MpJ superhealer mice enhances articular cartilage repair in a mouse injury model. *Stem Cell Res Ther*. 2020; 11: 93.
41. Zhou Y, Chyu J, Zumwalt M. Recent progress of fabrication of cell scaffold by electrospinning technique for articular cartilage tissue engineering. *Int J Biomater*. 2018; 2018: 1953636.
42. Hsieh-Bonassera ND, Wu I, Lin JK, Schumacher BL, Chen AC, Masuda K, et al. Expansion and redifferentiation of chondrocytes from osteoarthritic cartilage: cells for human cartilage tissue engineering. *Tissue Eng Part A*. 2009; 15: 3513-23.
43. Lee S, Kim JH, Jo CH, Seong SC, Lee JC, Lee MC. Effect of serum and growth factors on chondrogenic differentiation of synovium-derived stromal cells. *Tissue Eng Part A*. 2009; 15: 3401-15.
44. Hsieh-Bonassera ND, Wu I, Lin JK, Schumacher BL, Chen AC, Masuda K, et al. Expansion and redifferentiation of chondrocytes from osteoarthritic cartilage: cells for human cartilage tissue engineering. *Tissue Eng Part A*. 2009; 15: 3513-23.
45. Melero-Martin J, Al-Rubeai M. In vitro expansion of chondrocytes. *Top Tissue Eng*. 2007; 3: 37.
46. Malda J, Frondoza CG. Microcarriers in the engineering of cartilage and bone. *Trends Biotechnol*. 2006; 24: 299-304.
47. Zykwincka A, Marquis M, Godin M, Marchand L, Siquin C, Garnier C, et al. Microcarriers based on glycosaminoglycan-like marine exopolysaccharide for TGF- β 1 long-term protection. *Mar Drugs*. 2019; 17: 65.
48. Yang Y, Lin H, Shen H, Wang B, Lei G, Tuan RS. Mesenchymal stem cell-derived extracellular matrix enhances chondrogenic phenotype of and cartilage formation by encapsulated chondrocytes in vitro and in vivo. *Acta Biomater*. 2018; 69: 71-82.
49. Mithoefer K, Peterson L, Saris DB, Mandelbaum BR. Evolution and current role of autologous chondrocyte implantation for treatment of articular cartilage defects in the football (soccer) player. *Cartilage*. 2012; 3: 31S-6S.

50. Herberts CA, Kwa MS, Hermsen HP. Risk factors in the development of stem cell therapy. *J Transl Med.* 2011; 9: 29.
51. Kim N, Cho SG. New strategies for overcoming limitations of mesenchymal stem cell-based immune modulation. *Int J Stem Cells.* 2015; 8: 54-68.
52. Gimona M, Pachler K, Laner-Plamberger S, Schallmoser K, Rohde E. Manufacturing of human extracellular vesicle-based therapeutics for clinical use. *Int J Mol Sci.* 2017; 18: 1190.
53. Li X, Corbett AL, Taatizadeh E, Tasnim N, Little JP, Garnis C, et al. Challenges and opportunities in exosome research—perspectives from biology, engineering, and cancer therapy. *APL Bioeng.* 2019; 3: 011503.
54. Taghiyar L, Jahangir S, Khozaei Ravari M, Shamekhi MA, Eslaminejad MB. Cartilage repair by mesenchymal stem cell-derived exosomes: preclinical and clinical trial update and perspectives. In: Turksen K, editor. *Cell biology and translational medicine. Vol. 12: Stem Cells in Development and Disease.* Cham: Springer International Publishing. 2021; 1326: 73-93.
55. Zhang L, Hao C, Yao S, Tang R, Guo W, Cong H, et al. Exosomal miRNA profiling to identify nanoparticle phagocytic mechanisms. *Small.* 2018; 14: 1704008.
56. Kim DK, Kang B, Kim OY, Choi DS, Lee J, Kim SR, et al. EVpedia: an integrated database of high-throughput data for systemic analyses of extracellular vesicles. *J Extracell Vesicles.* 2013; 2: 20384.
57. Mao Y, Block T, Singh-Varma A, Sheldrake A, Leeth R, Griffey S, et al. Extracellular matrix derived from chondrocytes promotes rapid expansion of human primary chondrocytes in vitro with reduced dedifferentiation. *Acta Biomater.* 2019; 85: 75-83.
58. Cao Y, Vacanti JP, Paige KT, Upton J, Vacanti CA. Transplantation of chondrocytes utilizing a polymer-cell construct to produce tissue-engineered cartilage in the shape of a human ear. *Plast Reconstr Surg.* 1997; 100: 297-302.
59. Ma Y, Zhang W, Wang Z, Wang Z, Xie Q, Niu H, et al. Pegylated poly(glycerol sebacate)-modified calcium phosphate scaffolds with desirable mechanical behavior and enhanced osteogenic capacity. *Acta Biomater.* 2016; 44: 110-24.
60. Hessvik NP, Llorente A. Current knowledge on exosome biogenesis and release. *Cell Mol Life Sci.* 2018; 75: 193-208.
61. Doyle LM, Wang MZ. Overview of extracellular vesicles, their origin, composition, purpose, and methods for exosome isolation and analysis. *Cells.* 2019; 8.
62. Chen H, Wang L, Zeng X, Schwarz H, Nanda HS, Peng X, et al. Exosomes, a new star for targeted delivery. *Front Cell Dev Biol.* 2021; 9: 751079.
63. Xu L, Wu Y, Liu Y, Zhou Y, Ye Z, Tan WS. Non-contact coculture reveals a comprehensive response of chondrocytes induced by mesenchymal stem cells through trophic secretion. *Tissue Eng Regen Med.* 2018; 15: 37-48.
64. Shen Z, Huang W, Liu J, Tian J, Wang S, Rui K. Effects of mesenchymal stem cell-derived exosomes on autoimmune diseases. *Front Immunol.* 2021; 12: 749192.
65. Cosenza S, Ruiz M, Toupet K, Jorgensen C, Noël D. Mesenchymal stem cells derived exosomes and microparticles protect cartilage and bone from degradation in osteoarthritis. *Sci Rep.* 2017; 7: 16214.
66. Ni Z, Zhou S, Li S, Kuang L, Chen H, Luo X, et al. Exosomes: roles and therapeutic potential in osteoarthritis. *Bone Res.* 2020; 8: 25.
67. Newman CM, Bettinger T. Gene therapy progress and prospects: ultrasound for gene transfer. *Gene Ther.* 2007; 14: 465-75.
68. Mykhaylyk O, Antequera YS, Vlaskou D, Plank C. Generation of magnetic nonviral gene transfer agents and magnetofection in vitro. *Nat Protoc.* 2007; 2: 2391-411.
69. Evans CH, Ghivizzani SC, Robbins PD. Getting arthritis gene therapy into the clinic. *Nat Rev Rheumatol.* 2011; 7: 244-9.
70. Evans CH, Ghivizzani SC, Robbins PD. Gene therapy of the rheumatic diseases: 1998 to 2008. *Arthritis Res Ther.* 2009; 11: 209.
71. Evans CH, Ghivizzani SC, Robbins PD. Arthritis gene therapy's first death. *Arthritis Res Ther.* 2008; 10: 110.
72. Madry H, Orth P, Cucchiari M. Gene therapy for cartilage repair. *Cartilage.* 2011; 2: 201-25.
73. Harridge SD, Velloso CP. IGF-I and GH: potential use in gene doping. *Growth Horm IGF Res.* 2009; 19: 378-82.
74. Rodriguez-Merchan EC, Valentino LA. The role of gene therapy in cartilage repair. *Arch Bone Jt Surg.* 2019; 7: 79-90.
75. Gudas R, Simonaityte R, Riauba L, Pocius G, Kalesinskas R. [Treatment of joint surface pathology by Pridie drilling]. *Medicina (Kaunas).* 2002; 38: 720-9.
76. Fiorica C, Palumbo FS, Pitarresi G, Gulino A, Agnello S, Giammona G. Injectable in situ forming hydrogels based on natural and synthetic polymers for potential application in cartilage repair. *RSC Adv.* 2015; 5: 19715-23.
77. Dubruel P, Van Vlierberghe S. Biomaterials for bone regeneration: novel techniques and applications. Elsevier. 2014.
78. Elisseeff J, Anseth K, Sims D, McIntosh W, Randolph M, Yaremchuk M, et al. Transdermal photopolymerization of poly (ethylene oxide)-based injectable hydrogels for tissue-engineered cartilage. *Plast Reconstr Surg.* 1999; 104: 1014-22.
79. Nair LS. *Injectable hydrogels for regenerative engineering.* World Scientific. 2015.
80. Jeuken RM, Roth AK, Peters RJRW, Van Donkelaar CC, Thies JC, Van Rhijn LW, et al. Polymers in cartilage defect repair of the knee: current status and future prospects. *Polymers.* 2016; 8: 219.
81. Everhart JS, Jiang EX, Poland SG, Du A, Flanigan DC. Failures, reoperations, and improvement in knee symptoms following matrix-assisted autologous chondrocyte transplantation: a meta-analysis of prospective comparative trials. *Cartilage.* 2021; 13: 1022S-35S.
82. Pfeifer CG, Berner A, Koch M, Krutsch W, Kujat R, Angele P, et al. Higher ratios of hyaluronic acid enhance chondrogenic differentiation of human MSCs in a hyaluronic acid-gelatin composite scaffold. *Materials (Basel).* 2016; 9: 381.
83. Johnstone B, Alini M, Cucchiari M, Dodge GR, Eglin D, Guilak F, et al. Tissue engineering for articular cartilage repair—the state of the art. *Eur Cell Mater.* 2013; 25: 248-67.
84. Shi D, Xu X, Ye Y, Song K, Cheng Y, Di J, et al. Photo-cross-linked scaffold with kartogenin-encapsulated nanoparticles for cartilage regeneration. *ACS Nano.* 2016; 10: 1292-9.
85. Tharmalingam T, Sunley K, Spearman M, Butler M. Enhanced production of human recombinant proteins from CHO cells grown to high densities in macroporous microcarriers. *Mol Biotechnol.* 2011; 49: 263-76.
86. Leong W, Wang DA. Cell-laden polymeric microspheres for biomedical applications. *Trends Biotechnol.* 2015; 33: 653-66.

87. Sulaiman SB, Idrus RBH, Hwei NM. Gelatin microsphere for cartilage tissue engineering: current and future strategies. *Polymers*. 2020; 12: 2404.
88. Huang H, Xu H, Zhang J. Current tissue engineering approaches for cartilage regeneration. In: *Cartilage tissue engineering and regeneration techniques*. IntechOpen. 2019.
89. Lahner M, Duif C, Ficklscherer A, Kaps C, Kalwa L, Seidl T. Arthroscopic fixation of cell free polymer-based cartilage implants with a bioinspired polymer surface on the hip joint: a cadaveric pilot study. *BioMed Res Int*. 2014; 2014: 717912.
90. Yilmaz EN, Zeugolis DI. Electrospun polymers in cartilage engineering—state of play. *Front Bioeng Biotechnol*. 2020; 8: 77.
91. Guoyi L, et al. Establishing a tube foam scaffold for tracheal cartilage tissue engineering by using solvent casting/particulate leaching method. *Chin J Tissue Eng Res*. 2007; 11: 6278-81.
92. Chan BP, Leong KW. Scaffolding in tissue engineering: general approaches and tissue-specific considerations. *Eur Spine J*. 2008; 17: 467-79.
93. Di Bella C, Duchi S, O'Connell CD, Blanchard R, Augustine C, Yue Z, et al. In situ handheld three-dimensional bioprinting for cartilage regeneration. *J Tissue Eng Regen Med*. 2018; 12: 611-21.
94. Hurst JM, Steadman JR, O'Brien L, Rodkey WG, Briggs KK. Rehabilitation following microfracture for chondral injury in the knee. *Clin Sports Med*. 2010; 29: 257-65.
95. Miller BS, Steadman JR, Briggs KK, Rodrigo JJ, Rodkey WG. Patient satisfaction and outcome after microfracture of the degenerative knee. *J Knee Surg*. 2004; 17: 13-7.
96. Kreuz PC, Erggelet C, Steinwachs MR, Krause SJ, Lahm A, Niemeyer P, et al. Is microfracture of chondral defects in the knee associated with different results in patients aged 40 years or younger? *Arthroscopy*. 2006; 22: 1180-6.
97. Erggelet C, Endres M, Neumann K, Morawietz L, Ringe J, Haberstroh K, et al. Formation of cartilage repair tissue in articular cartilage defects pretreated with microfracture and covered with cell-free polymer-based implants. *J Orthop Res*. 2009; 27: 1353-60.
98. Yang HS, La WG, Bhang SH, Kim HJ, Im GI, Lee H, et al. Hyaline cartilage regeneration by combined therapy of microfracture and long-term bone morphogenetic protein-2 delivery. *Tissue Eng Part A*. 2011; 17: 1809-18.
99. Guo T, Noshin M, Baker HB, Taskoy E, Meredith SJ, Tang Q, et al. 3D printed biofunctionalized scaffolds for microfracture repair of cartilage defects. *Biomaterials*. 2018; 185: 219-31.
100. Hangody L, Kish G, Kárpáti Z, Szerb I, Udvarhelyi I. Arthroscopic autogenous osteochondral mosaicplasty for the treatment of femoral condylar articular defects A preliminary report. *Knee Surg Sports Traumatol Arthrosc*. 1997; 5: 262-7.
101. Cognault J, Seurat O, Chaussard C, Ionescu S, Saragaglia D. Return to sports after autogenous osteochondral mosaicplasty of the femoral condyles: 25 cases at a mean follow-up of 9 years. *Orthop Traumatol Surg Res*. 2015; 101: 313-7.
102. Chuang E-Y, Chiang C, Wong P, Chen C. Hydrogels for the application of articular cartilage tissue Engineering: a review of hydrogels. *Adv Mater Sci Eng*. 2018; 2018: 1-13.
103. Bentley G, Biant LC, Vijayan S, Macmull S, Skinner JA, Carrington RW. Minimum ten-year results of a prospective randomised study of autologous chondrocyte implantation versus mosaicplasty for symptomatic articular cartilage lesions of the knee. *J Bone Joint Surg Br*. 2012; 94: 504-9.
104. Bartha L, Hamann D, Pieper J, Péters F, Riesle J, Vajda A, et al. A clinical feasibility study to evaluate the safety and efficacy of PEOT/PBT implants for human donor site filling during mosaicplasty. *Eur J Orthop Surg Traumatol*. 2013; 23: 81-91.
105. Battaglia M, Rimondi E, Monti C, Guaraldi F, Sant'Andrea A, Buda R, et al. Validity of T2 mapping in characterization of the regeneration tissue by bone marrow derived cell transplantation in osteochondral lesions of the ankle. *Eur J Radiol*. 2011; 80: e132-9.
106. Buda R, Vannini F, Cavallo M, Baldassarri M, Luciani D, Mazzotti A, et al. One-step arthroscopic technique for the treatment of osteochondral lesions of the knee with bone-marrow-derived cells: three years results. *Musculoskelet Surg*. 2013; 97: 145-51.
107. Giannini S, Buda R, Battaglia M, Cavallo M, Ruffilli A, Ramponi L, et al. One-step repair in talar osteochondral lesions: 4-year clinical results and t2-mapping capability in outcome prediction. *Am J Sports Med*. 2013; 41: 511-8.
108. Woodmass JM, Melugin HP, Wu IT, Saris DBF, Stuart MJ, Krych AJ. Viable osteochondral allograft for the treatment of a full-thickness cartilage defect of the patella. *Arthrosc Tech*. 2017; 6: e1661-5-e1665.
109. Melugin HP, Ridley TJ, Bernard CD, Wischmeier D, Farr J, Stuart MJ, et al. Prospective outcomes of cryopreserved osteochondral allograft for patellofemoral cartilage defects at minimum 2-year follow-up. *Cartilage*. 2021; 13: 1014S-21S.
110. Janis L, Kaplansky DB, DeCarbo WT. Early clinical experience with a fresh talar transplant inlay allograft for the treatment of osteochondral lesions of the talus. *J Am Podiatr Med Assoc*. 2010; 100: 25-34.
111. Scranton Jr PE, Frey CC, Feder KS. Outcome of osteochondral autograft transplantation for type-V cystic osteochondral lesions of the talus. *J Bone Joint Surg Br*. 2006; 88: 614-9.
112. Scranton Jr PE, McDermott JE. Treatment of type V osteochondral lesions of the talus with ipsilateral knee osteochondral autografts. *Foot Ankle Int*. 2001; 22: 380-4.
113. Heesch KC. Increased physical activity in older women could reduce the burden of arthritis. *Nature Reviews Rheumatology*. 2007; 3: 371.
114. Siclari A, Mascaro G, Kaps C, Boux E. A 5-year follow-up after cartilage repair in the knee using a platelet-rich plasma-immersed polymer-based implant. *Open Orthop J*. 2014; 8: 346-54.
115. Yamaguchi Jr KT, Mosich GM, Jones KJ. Arthroscopic delivery of injectable bone graft for staged revision anterior cruciate ligament reconstruction. *Arthrosc Tech*. 2017; 6: e2223-7-e2227.
116. Ng J, Spiller K, Bernhard J, Vunjak-Novakovic G. Biomimetic approaches for bone tissue engineering. *Tissue Eng Part B Rev*. 2017; 23: 480-93.
117. Niethammer TR, Pietschmann MF, Horng A, Roßbach BP, Ficklscherer A, Jansson V, et al. Graft hypertrophy of matrix-based autologous chondrocyte implantation: a two-year follow-up study of NOVOCART 3D implantation in the knee. *Knee Surg Sports Traumatol Arthrosc*. 2014; 22: 1329-36.
118. Villalba J, Peñalver JM, Sánchez J. Tratamiento de los grandes defectos osteocondrales de cóndilo femoral externo en pacientes jóvenes mediante autoinjerto y malla de colágeno. *Rev Española Cir Ortop Traumatol*. 2021; 65: 317-21.
119. Stoker AM, Stannard JP, Cook JL. Chondrocyte viability at time of transplantation for osteochondral allografts preserved by the Missouri Osteochondral Preservation System versus standard tissue bank protocol. *J Knee Surg*. 2018; 31: 772-80.

120. Stannard JP, Cook JL. Prospective assessment of outcomes after primary unipolar, multisurface, and bipolar osteochondral allograft transplantations in the knee: a comparison of 2 preservation methods. *Am J Sports Med.* 2020; 48: 1356-64.
121. Siclari A, Mascaro G, Gentili C, Cancedda R, Boux E. A cell-free scaffold-based cartilage repair provides improved function hyaline-like repair at one year. *Clin Orthop Relat Res.* 2012; 470: 910-9.
122. Gerlier L, Lamotte M, Wille M, Kreuz PC, Vanlauwe J, Dubois D, et al. The cost utility of autologous chondrocytes implantation using ChondroCelect® in symptomatic knee cartilage lesions in Belgium. *Pharmacoeconomics.* 2010; 28: 1129-46.
123. Syed BA, Evans JB. Stem cell therapy market. *Nat Rev Drug Discov.* 2013; 12: 185-6.
124. Jones HR, Crawford DC. An autologous tissue implant, NeoCart, for treatment of hyaline cartilage injury in the knee. *Oper Tech Orthop.* 2014; 24: 264-70.
125. Ryan Delaney M, Barrett BC, Stevens MP. ProChondrix® cartilage restoration matrix contains growth factors necessary for hyaline cartilage regeneration. 2016.
126. Shive MS, Stanish WD, McCormack R, Forriol F, Mohtadi N, Pelet S, et al. BST-CarGel® treatment maintains cartilage repair superiority over microfracture at 5 years in a multicenter randomized controlled trial. *Cartilage.* 2015; 6: 62-72.
127. Hoemann CD, Sun J, McKee MD, Chevri er A, Rossomacha E, Rivard GE, et al. Chitosan-glycerol phosphate/blood implants elicit hyaline cartilage repair integrated with porous subchondral bone in microdrilled rabbit defects. *Osteoarthr Cartil.* 2007; 15: 78-89.
128. Carey JL, Remmers AE, Flanigan DC. Use of MACI (autologous cultured chondrocytes on porcine collagen membrane) in the United States: preliminary experience. *Orthop J Sports Med.* 2020; 8: 2325967120941816.
129. Gomoll AH. Osteochondral allograft transplantation using the chondrofix implant. *Oper Tech Sports Med.* 2013; 21: 90-4.
130. Buckwalter JA, Bowman GN, Albright JP, Wolf BR, Bollier M. Clinical outcomes of patellar chondral lesions treated with juvenile particulated cartilage allografts. *Iowa Orthop J.* 2014; 34: 44-9.
131. Blunt L, Bills PJ, Fleming L. Metrology for bio systems; 2015.
132. Chubinskaya S, Di Matteo B, Lovato L, Iacono F, Robinson D, Kon E. Agili-C implant promotes the regenerative capacity of articular cartilage defects in an ex vivo model. *Knee Surg Sports Traumatol Arthrosc.* 2019; 27: 1953-64.
133. Scarritt M, Murdock M, Badylak SF. Biologic scaffolds composed of extracellular matrix for regenerative medicine. In: *Principles of regenerative medicine.* Elsevier. 2019; 613-26.
134. Hinckel BB, Thomas D, Vellios EE, Hancock KJ, Calcei JG, Sherman SL, et al. Algorithm for treatment of focal cartilage defects of the knee: classic and new procedures. *Cartilage.* 2021; 13: 473S-95S.
135. Younesi M, Goldberg VM, Akkus O. A micro-architecturally biomimetic collagen template for mesenchymal condensation based cartilage regeneration. *Acta Biomater.* 2016; 30: 212-21.
136. Stack JD, Levingstone TJ, Lalor W, Sanders R, Kearney C, O'Brien FJ, et al. Repair of large osteochondritis Dissecans lesions using a novel multilayered tissue engineered construct in an equine athlete. *J Tissue Eng Regen Med.* 2017; 11: 2785-95.
137. Goodfriend B, Essilfie AA, Jones IA, Thomas Vangsness C. Fresh osteochondral grafting in the United States: the current status of tissue banking processing. *Cell Tissue Banking.* 2019; 20: 331-7.
138. Rundell VL, Beck RT, Wang CE, Gutowski KA, Sisco M, Fenner G, et al. Complication prevalence following use of tutoplast-derived human acellular dermal matrix in prosthetic breast reconstruction: A retrospective review of 203 patients. *J Plast Reconstr Aesthet Surg.* 2014; 671: 345-51.
139. Bastian JD, B uchler L, Meyer DC, Siebenrock KA, Keel MJ. Surgical hip dislocation for osteochondral transplantation as a salvage procedure for a femoral head impaction fracture. *J Orthop Trauma.* 2010; 24: e113-8.
140. Holmes B, et al. Testing of a 3D printed, nanostructured osteochondral implant for knee repair in a small animal model. 2017.
141. Jeznach O, Kołbuk D, Sajkiewicz P. Injectable hydrogels and nanocomposite hydrogels for cartilage regeneration. *J Biomed Mater Res A.* 2018; 106: 2762-76.
142. Cucchiari ni M, Madry H, Guilak F, Saris DB, Stoddart MJ, Koon Wong M, et al. A vision on the future of articular cartilage repair. *Eur Cell Mater.* 2014; 27: 12-6.
143. Spalding T. Cartilage repair: autograft osteochondral grafting. In: *Arthroscopy.* Springer. 2016; 173-87.
144. Whitehouse MR, Howells NR, Parry MC, Austin E, Kafienah W, Brady K, et al. Repair of torn avascular meniscal cartilage using undifferentiated autologous mesenchymal stem cells: from in vitro optimization to a first-in-human study. *Stem Cells Transl Med.* 2017; 6: 1237-48.
145. Barlic A, et al. Advancing in the quality of the cells assigned for autologous chondrocyte implantation (ACI) method. In: *11th Mediterranean Conference on Medical and Biomedical Engineering and Computing 2007.* Springer. 2007.
146. Arabpour Z, Baradaran-Rafii A, Bakhshaiesh NL, Ai J, Ebrahimi-Barough S, Esmaeili Malekabadi H, et al. Design and characterization of biodegradable multi layered electrospun nanofibers for corneal tissue engineering applications. *J Biomed Mater Res A.* 2019; 107: 2340-9.
147. Aggarwal S, Pittenger MF. Human mesenchymal stem cells modulate allogeneic immune cell responses. *Blood.* 2005; 105: 1815-22.
148. Engela AU, Hoogduijn MJ, Boer K, Litjens NH, Betjes MG, Weimar W, et al. Human adipose-tissue derived mesenchymal stem cells induce functional de-novo regulatory T cells with methylated FOXP3 gene DNA. *Clin Exp Immunol.* 2013; 173: 343-54.
149. Maggini J, Mirkin G, Bognanni I, Holmberg J, Piazz on IM, Nepomnaschy I, et al. Mouse bone marrow-derived mesenchymal stromal cells turn activated macrophages into a regulatory-like profile. *PLOS ONE.* 2010; 5: e9252.
150. GBD 2017 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet.* 2018; 392: 1789-858.
151. Simorgh S, Milan PB, Saadatmand M, Bagher Z, Gholipourmalekabadi M, Alizadeh R, et al. Human olfactory mucosa stem cells delivery using a collagen hydrogel: as a potential candidate for bone tissue engineering. *Materials (Basel).* 2021; 14: 3909.
152. Je JY, Kim SK. Reactive oxygen species scavenging activity of aminoderivatized chitosan with different degree of deacetylation. *Bioorg Med Chem.* 2006; 14: 5989-94.
153. Hirabara S, Kojima T, Takahashi N, Hanabayashi M, Ishiguro N. Hyaluronan inhibits TLR-4 dependent cathepsin K and matrix metalloproteinase 1 expression in human fibroblasts. *Biochem Biophys Res Commun.* 2013; 430: 519-22.

154. Swartzlander MD, Blakney AK, Amer LD, Hankenson KD, Kyriakides TR, Bryant SJ. Immunomodulation by mesenchymal stem cells combats the foreign body response to cell-laden synthetic hydrogels. *Biomaterials*. 2015; 41: 79-88.
155. Veiseh O, Doloff JC, Ma M, Vegas AJ, Tam HH, Bader AR, et al. Size- and shape-dependent foreign body immune response to materials implanted in rodents and nonhuman primates. *Nat Mater*. 2015; 14: 643-51.
156. Vallés G, Bensiamar F, Crespo L, Arruebo M, Vilaboa N, Saldaña L. Topographical cues regulate the crosstalk between MSCs and macrophages. *Biomaterials*. 2015; 37: 124-33.
157. Yang F, Zhang Y, Liu B, Cao M, Yang J, Tian F, et al. Basic fibroblast growth factor and agarose gel promote the ability of immune privilege of allogeneic cartilage transplantation in rats. *J Orthop Translat*. 2020; 22: 73-80.
158. Hori S, Nomura T, Sakaguchi S. Control of regulatory T cell development by the transcription factor Foxp3. *Science*. 2003; 299: 1057-61.
159. Zhang J, Huang X, Wang H, Liu X, Zhang T, Wang Y, et al. The challenges and promises of allogeneic mesenchymal stem cells for use as a cell-based therapy. *Stem Cell Res Ther*. 2015; 6: 234.
160. Almqvist KF, Dhollander AA, Verdonk PC, Forsyth R, Verdonk R, Verbruggen G. Treatment of cartilage defects in the knee using alginate beads containing human mature allogenic chondrocytes. *Am J Sports Med*. 2009; 37: 1920-9.
161. de la Garza-Rodea AS, Verweij MC, Boersma H, van der Velde-van Dijke I, de Vries AA, Hoeben RC, et al. Exploitation of herpesvirus immune evasion strategies to modify the immunogenicity of human mesenchymal stem cell transplants. *PLOS ONE*. 2011; 6: e14493.
162. Sullivan C, Barry F, Ritter T, O'Flatharta C, Howard L, Shaw G, et al. Allogeneic murine mesenchymal stem cells: migration to inflamed joints in vivo and amelioration of collagen induced arthritis when transduced to express CTLA4lg. *Stem Cells Dev*. 2013; 22: 3203-13.
163. Chen W, Li M, Su G, Zang Y, Yan Z, Cheng H, et al. Co-transplantation of hematopoietic stem cells and cxcr4 gene-transduced mesenchymal stem cells promotes hematopoiesis. *Cell Biochem Biophys*. 2015; 71: 1579-87.
164. Heldens GT, Blaney Davidson EN, Vitters EL, Schreurs BW, Piek E, van den Berg WB, et al. Catabolic factors and osteoarthritis-conditioned medium inhibit chondrogenesis of human mesenchymal stem cells. *Tissue Eng Part A*. 2012; 18: 45-54.
165. Khan IM, Gilbert SJ, Singhrao SK, Duance VC, Archer CW. Cartilage integration: evaluation of the reasons for failure of integration during cartilage repair. A review. *Eur Cell Mater*. 2008; 16: 26-39.
166. Montaseri A, et al. IGF-1 and PDGF-bb suppress IL-1 β -induced cartilage degradation through down-regulation of NF- κ B signaling: involvement of Src/PI-3K/AKT pathway. *PLOS ONE*. 2011; 6: 1-15.
167. Rahmani Del Bakhshayesh A, Babaie S, Tayefi Nasrabadi H, Asadi N, Akbarzadeh A, Abedelahi A. An overview of various treatment strategies, especially tissue engineering for damaged articular cartilage. *Artif Cells Nanomed Biotechnol*. 2020; 48: 1089-104.
168. Braun HJ, Kim HJ, Chu CR, Dragoo JL. The effect of platelet-rich plasma formulations and blood products on human synovio-cytes: implications for intra-articular injury and therapy. *Am J Sports Med*. 2014; 42: 1204-10.
169. Sánchez M, Delgado D, Garate A, Sánchez P, Padilla S, Azofra J. Platelet-rich plasma combined with allograft to treat osteochondritis Dissecans of the knee: a case report. *J Med Case Rep*. 2019; 13: 105.
170. Sundman EA, Cole BJ, Karas V, Della Valle C, Tetreault MW, Mohammed HO, et al. The anti-inflammatory and matrix restorative mechanisms of platelet-rich plasma in osteoarthritis. *Am J Sports Med*. 2014; 42: 35-41.
171. Bendinelli P, Matteucci E, Dogliotti G, Corsi MM, Banfi G, Maroni P, et al. Molecular basis of anti-inflammatory action of platelet-rich plasma on human chondrocytes: mechanisms of NF- κ B inhibition via HGF. *J Cell Physiol*. 2010; 225: 757-66.